

Bile Acids as Receptor Ligands in Metabolic Processes - The FXR Connection

Sarah Louise Long^{1,2} and Susan A. Joyce^{1,2*}

¹School of Biochemistry and Cell Biology, University College Cork, Cork, Ireland.

²APC Microbiome Ireland, University College Cork, Cork, Ireland

Introduction

Cell signaling in response to diet and metabolites are important mediators for metabolic outcomes. They are different in the fed and fasted states. In recent years postprandial Bile Acids (BAs), oxysterols, hormones and indeed fatty acids (FAs) that are derived from dietary and human lipids including cholesterol derivatives or its precursors are recognised as differential signaling molecules to interact with an array of two distinct classes of receptors that are central to metabolism: nuclear receptors and G-Protein Coupled Receptors (GPCRs). Two main BA modulated receptors, nuclear receptor Farnesoid X Receptor (FXR) and Takeda G protein coupled receptor 5 (TGR5), are central to lipid and carbohydrate metabolism and in cross talk between the gut to the liver and with other tissues. They also influence the action of other NRs and GPCRs. Table 1 summarizes the characterized BA activated receptors, their ligand agonists, the tissues where they are expressed and the processes that they influence. This review focuses on the FXR, its biochemistry, functions and implications for health and in disease.

Nuclear receptors (NRs) including FXR, are cell membrane, ligand activated, transcription factors with an important role in regulating many physiological pathways within a cell [1] (Table 1). Once activated NRs move to the cell nucleus to alter the transcriptional landscape by binding to recognition elements (REs) in target gene promoter/s [2]. NRs can act as homodimers or heterodimers and can bind to a variety of DNA response elements that are specific for each NR [3]. They influence a range of metabolic processes and this is reflected by their range of ligands (Table 1). Note, that there is considerable overlap among ligands, but that they vary in their relative potency for receptor activation. Importantly, therefore, they appear to cross talk for efficiency and sometimes amplification of function.

FXR, known as the BA receptor (BAR), since it is potently activated by individual BA moieties, it controls BA levels but it is also central to lipid and sugar metabolism in both the liver and the gut. BAs are central for micelle formation and nutrient uptake. GIT FXR is postprandially activated in response to gall bladder BA release and the moieties generated by GIT microbial action [4]. In the liver, recycled BA concentrations alter its expression to regulate the synthesis of BA, and in doing so, alter cholesterol, lipid, and glucose as well as autophagy-mediated lipid catabolism [5-8]. A further layer of regulation is through gut to liver hormone FGF19, shutting off the rate-limiting step in bile acid synthesis through CYP7A1. Its phosphorylation in hepatocytes facilitates its transport to the nucleus to activate non-receptor tyrosine kinase (Src) phosphorylation to further activate FXR (on Y67) [9]. Indeed, these authors report that FXR phosphorylation and signaling appears defective in primary biliary cirrhosis (PBC) sufferers.

While two FXR alleles are present in humans, designated FXR α (NR1H4) and FXR β (NR1H5) [10], FXR β is a pseudogene in humans

Corresponding Author: Joyce SA, School of Biochemistry and Cell Biology, APC Microbiome Ireland, University College Cork, Cork, Ireland. E-mail id: S.Joyce@ucc.ie

Received Date: Nov 29, 2019;

Accepted Date: Dec 06, 2019;

Published Date: Dec 11, 2019

Publisher: Scholars Insight Online Publishers

Citation: Long SL, Joyce SA. Bile Acids as Receptor Ligands in Metabolic Processes - The FXR Connection. J Food Nutr Sci Int. 2019; 1:101.

Copyright: ©2019 Long SL. This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

but is active in rats and rabbits [11]. FXR α is conserved across many species from human to fish [12]. Alternative RNA processing identifies two distinct promoters but four different isoforms of this protein. Isoforms 1 and 2 represent the active forms [10,13]. FXR α is differentially expressed in tissues including liver, intestine, kidney and adrenal gland, while low expression is detected in the heart, adipose tissue and skeletal muscle [3,14]. Thus, we can hypothesize that FXR is important for frontline energetics and less so for stored energy. Indeed, FXR is also an important regulator of lipid and glucose metabolism. In the diabetic murine model FXR activation leads to reduced circulating glucose concentrations improving hyperglycaemia [15]. An effect later confirmed in FXR knockout mice [16]. Synthetic FXR agonist fexeramine (FEX), reduces adiposity and weight gain while improving insulin sensitivity, cholesterol levels and reducing the levels of inflammatory cytokines in a DIO mouse model [17,18]. Activation of FXR can also reduce inflammation, seen in cases of diabetic nephropathy [19]. Since FXR activation displays beneficial effects to various metabolic associated diseases, this receptor is targeted for metabolic treatments [20].

The liver has a role in controlling plasma glucose homeostasis through the activation and expression of genes encoding metabolic enzymes including glucokinase, L-type Pyruvate Kinase (Lpk) and Acetyl-coenzymeA carboxylase-1 (Acc-1) mediated by the sterol regulatory element-binding protein-1c (Srebp-1c) and carbohydrate-response elements (ChoREs) [21]. These enzymes maintain a balance between glucose uptake and production during fed and fast states in response to FXR signaling [15,22]. FXR also regulates triglyceride production in the liver. Here, FXR activation, by Cholic Acid (CA), suppresses expression of the transcription factor SREBP-1c causing

Table 1: Bile acid activated nuclear receptors. The table includes endogenous bile acid ligands, site of action, mechanism of action and downstream effects.

Nuclear Receptor	Cofactors	Bile acid agonists	Site of action	Process	References
FXR	DRIP205, SRC,	CDCA>LCA=DCA>CA	Brain, lungs, liver, kidney, adrenal gland	Regulate glucose and lipid metabolism, ↓inflammation	(Pineda Torra et al., 2004, Bramlett et al., 2000)
RXR	PGC-1, TIF2	Retinoic acid, 7-Hydroxycholesterol	Brain, lungs, liver, kidney, intestine	Maintain glucose and lipid metabolism, bone development, immunity,	(Delerive et al., 2002, Dawson and Xia 2012, Menendez-Gutierrez and Ricote 2017)
LXR	SIRT1, p160, TRRAP	Oxysterols, 27-hydroxycholesterol, 6 α -hydroxy bile acid (HDCA)	Brain, lungs, liver, kidney, intestine, adipose tissue, adrenal gland, spleen	↑cholesterol metabolism and efflux	(Bovenge et al., 2015, Husskonen et al., 2004, Unno et al., 2005)
PXR	PGC-1 α , SGK2, PP2C α , SCR1/ NCOA1, SCR2/ GRIPI	3-KetolCA>LCA.DCA. CA	Intestine, liver	↑detoxification of BA, ↑dysregulation of lipid and glucose metabolism	(Wagner et al., 2005, Hakkola et al., 2016, Hassani-Nezhad-Gashti et al., 2018, Hariprasad et al., 2009)
VDR	SRC-1/TIF2, CBP/ p300, SRA/DRIP/ TRAP	3-oxo-LCA>LCA>DCA>CA	Bone, liver, intestines, kidneys, muscle, brain, and skin	↑detoxification of LCA ↑inflammation	(Freedman 1999, Makishima, Lu et al. 2002, Pols, Puchner et al. 2017)
PPAR	PBP/PPARBP, TRAP220/MED1, PGC-1 α , SRC-2/ TIF2/GRIPI	Essential Fatty acids, Trans-retinoic acid, eicosanoids	Liver, intestines, adipose tissue, heart, spleen and skeletal muscle	↑inflammation ↑fatty acid oxidation, maintain energy homeostasis	(Viswakarma et al., 2010, Graham et al., 2005, Wang et al., 2003, Wang et al., 2018)

down regulation of enzymes, including Fatty Acid Synthase (FAS), required for lipogenesis [23]. This FXR mediated inhibition of SREBP-1c expression involves interaction with Small Heterodimer Partner (SHP) and interference of Liver X Receptor (LXR) activity [23]. This FXR mediated SREBP-1c repression could prevent lipid synthesis and storage and could alter cholesterol synthesis again reinforcing FXR as a potential therapeutic target [24,25]. FXR mediated regulation of triglyceride levels can also be controlled by coactivator peroxisome-proliferator-activated receptor- γ -coactivator-1 α (PGC-1 α). PGC-1 α upregulates FXR expression by co-activation of peroxisome proliferator-activated receptor (PPAR γ) and HNF4 α to triglyceride synthesis and secretion [26]. FXR also dictates the flux of lipoproteins carrying lipid back to the liver for processing through activation of a number of surface receptors for low and very low density lipoprotein uptake. It can also determine the level of apolipoprotein levels to tag these and high density lipoproteins that dictate reverse cholesterol transport [27-35]. Taken together activation of FXR [36-40], whether it is phosphorylated or not, may influence different outcomes in different tissues and these outcomes appear dependent on the fed or the fasted state and enterohepatic circulation of dietary defendant hormones and metabolites from the gut [41-44].

Perspective

NRcrossstalk and their responses are modulated in the fed and in the fasted states, in doing so they can alter metabolism and influence immune function. Their relative expression in sustaining health and in contributing to diseases remains unknown and therefore merits further investigation through assessment of their combined therapeutic value.

Acknowledgements

SSL and SAJ are supported by SFI-EU JPI CABALA (Grant Number 16/ERA-HDHL/3358) and by Science Foundation of Ireland Centres for Science, Engineering and Technology (CSET) programme (Grant Number SFI/12/RC/2273) to APC Microbiome Ireland. SAJ is also supported by DAFM 17-RD-US-ROI.

References

- Huang P, Chandra V, Rastinejad F. "Structural Overview of the Nuclear Receptor Superfamily: Insights into Physiology and Therapeutics." Annual review of physiology. 2010; 72: 247-272.
- Bain DL, Heneghan AF, Connaghan-Jones KD, Miura MT. "Nuclear Receptor Structure: Implications for Function." Annual Review of Physiology. 2007; 69: 201-220.
- Zhang Y, Kast-Woelbern HR, Edwards PA. "Natural Structural Variants of the Nuclear Receptor Farnesoid X Receptor Affect Transcriptional Activation." Journal of Biological Chemistry. 2003; 278: 104-110.
- Long SL, Gahan CG, Joyce SA. "Interactions between gut bacteria and bile in health and disease." Molecular aspects of medicine. 2017; 56: 54-65.
- Lefebvre P, Cariou B, Lien F, Kuipers F, Staels B. "Role of Bile Acids and Bile Acid Receptors in Metabolic Regulation." Physiological Reviews. 2009; 89: 147-191.
- de Aguiar Vallim TQ, Tarling EJ, Edwards PA. "Pleiotropic roles of bile acids in metabolism." Cell Metab. 2013; 17: 657-669.
- Lee JM, Wagner M, Xiao R, Kim KH, Feng D, Lazar MA, et al. "Nutrient Sensing Nuclear Receptors Coordinate Autophagy." Nature. 2014; 516: 112-115.
- Seok S, Fu T, Choi SE, Li Y, Zhu R, Kumar S, et al. "Transcriptional regulation of autophagy by an FXR/CREB axis." Nature. 2014; 516: 108-111.
- Byun S, Kim DH, Ryerson D, Kim YC, Sun H, Kong B, et al. "Postprandial FGF19-induced phosphorylation by Src is critical for FXR function in bile acid homeostasis." Nature Communications. 2018; 9: 2590.
- Lee FY, Lee H, Hubbert ML, Edwards PA, Zhang Y. "FXR, a multipurpose nuclear receptor." Trends in Biochemical Sciences. 2006; 31: 572-580.

11. Otte KH, Kober KI, Thompson P, Hoefler M, Haubold B, Rimmel B, et al. "Identification of Farnesoid X Receptor β as a Novel Mammalian Nuclear Receptor Sensing Lanosterol." *Molecular and Cellular Biology*. 2003; 23: 864-872.
12. Han C. "Update on FXR Biology: Promising Therapeutic Target?" *International Journal of Molecular Sciences*. 2018; 19: 2069.
13. Vaquero J, Monte MJ, Dominguez M, Muntané J, Marin JGG. "Differential activation of the human farnesoid X receptor depends on the pattern of expressed isoforms and the bile acid pool composition." *Biochemical Pharmacology*. 2013; 86: 926-939.
14. Huber RM, Murphy K, Miao B, Link JR, Cunningham MR, Rupar MJ, et al. "Generation of multiple farnesoid-X-receptor isoforms through the use of alternative promoters." *Gene*. 2002; 290: 35-43.
15. Zhang Y, Lee FY, Barrera G, Lee H, Vales C, Gonzalez FJ, et al. "Activation of the nuclear receptor FXR improves hyperglycemia and hyperlipidemia in diabetic mice." *Proceedings of the National Academy of Sciences of the United States of America*. 2006; 103: 1006-1011.
16. Ma K, Saha PK, Chan L, Moore DD. "Farnesoid X receptor is essential for normal glucose homeostasis." *J Clin Invest*. 2006; 116: 1102-1109.
17. Downes M, Verdecia MA, Roecker AJ, Hughes R, Hogenesch JB, Kast-Woelbern HR, et al. "A Chemical, Genetic, and Structural Analysis of the Nuclear Bile Acid Receptor FXR." *Molecular Cell*. 2003; 11: 1079-1092.
18. Fang S, Suh JM, Reilly SM, Yu E, Osborn O, Lackey D, et al. "Intestinal FXR agonism promotes adipose tissue browning and reduces obesity and insulin resistance." *Nat Med*. 2015; 21: 159-165.
19. Herman-Edelstein M, Weinstein T, Levi M. "Bile acid receptors and the kidney." *Curr Opin Nephrol Hypertens*. 2018; 27: 56-62.
20. Chávez-Talavera O, Tailleux A, Lefebvre P, Staels B. "Bile Acid Control of Metabolism and Inflammation in Obesity, Type 2 Diabetes, Dyslipidemia, and Nonalcoholic Fatty Liver Disease." *Gastroenterology*. 2017; 152: 1679-1694.e1673.
21. Poupeau A, Postic C. "Cross-regulation of hepatic glucose metabolism via ChREBP and nuclear receptors." *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. 2011; 1812: 995-1006.
22. Duran-Sandoval D, Cariou B, Percevault F, Hennuyer N, Grefhorst A, van Dijk TH, et al. "The Farnesoid X Receptor Modulates Hepatic Carbohydrate Metabolism during the Fasting-Refeeding Transition." *Journal of Biological Chemistry*. 2005; 280: 29971-29979.
23. Watanabe M, Houten SM, Wang L, Moschetta A, Mangelsdorf DJ, Heyman RA, et al. "Bile acids lower triglyceride levels via a pathway involving FXR, SHP, and SREBP-1c." *Journal of Clinical Investigation*. 2004; 113: 1408-1418.
24. Karagianni P, Talianidis I. "Transcription factor networks regulating hepatic fatty acid metabolism." *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*. 2015; 1851: 2-8.
25. Liu N, Zhao J, Wang J, Teng H, Fu Y, Yuan H. "Farnesoid X receptor ligand CDCA suppresses human prostate cancer cells growth by inhibiting lipid metabolism via targeting sterol response element binding protein 1." *American Journal of Translational Research*. 2016; 8: 5118-5124.
26. Zhang Y, Castellani LW, Sinal CJ, Gonzalez FJ, Edwards PA. "Peroxisome proliferator-activated receptor- γ coactivator 1 α (PGC-1 α) regulates triglyceride metabolism by activation of the nuclear receptor FXR." *Genes & Development*. 2004; 18: 157-169.
27. Cariou B, Staels B. "FXR: a promising target for the metabolic syndrome?" *Trends Pharmacol Sci*. 2007; 28: 236-243.
28. Dawson MI, Xia Z. "The Retinoid X Receptors and Their Ligands." *Biochimica et biophysica acta*. 2012; 821: 21-56.
29. Delerive P, Wu Y, Burriss TP, Chin WW, Suen CS. "PGC-1 Functions as a Transcriptional Coactivator for the Retinoid X Receptors." *Journal of Biological Chemistry*. 2002; 277: 3913-3917.
30. Freedman LP. "Increasing the complexity of coactivation in nuclear receptor signaling." *Cell*. 1999; 97: 5-8.
31. Graham TL, Mookherjee C, Suckling KE, Palmer CAN, Patel L. "The PPAR δ agonist GW0742X reduces atherosclerosis in LDLR-/- mice." *Atherosclerosis*. 2005; 181: 29-37.
32. Hakkola J, Rysä J, Hukkanen J. "Regulation of hepatic energy metabolism by the nuclear receptor PXR." *Biochimica et Biophysica Acta (BBA) - Gene Regulatory Mechanisms*. 2016; 1859: 1072-1082.
33. Hariparsad N, Chu X, Yabut J, Labhart P, Hartley DP, Dai X, et al. "Identification of pregnane-X receptor target genes and coactivator and corepressor binding to promoter elements in human hepatocytes." *Nucleic Acids Research*. 2009; 37: 1160-1173.
34. Hassani-Nezhad-Gashti F, Rysä J, Kumm O, Nöpänkangas J, Buler M, Karpale M, et al. "Activation of nuclear receptor PXR impairs glucose tolerance and dysregulates GLUT2 expression and subcellular localization in liver." *Biochemical Pharmacology*. 2018; 148: 253-264.
35. Huuskonen J, Fielding PE, Fielding CJ. Role of p160 coactivator complex in the activation of liver X receptor. *Arterioscler Thromb Vasc Biol*. 2004; 24: 703-708.
36. Makishima M, Lu TT, Xie W, Whitfield GK, Domoto H, et al. Vitamin D Receptor As an Intestinal Bile Acid Sensor. *Science*. 2002; 29: 61313-61316.
37. Menéndez-Gutiérrez MP, Ricote M. The multi-faceted role of retinoid X receptor in bone remodeling. *Cellular and Molecular Life Sciences*. 2017; 74: 2135-2149.
38. Pineda Torra I, Freedman LP, Garabedian MJ. Identification of DRIP205 as a coactivator for the Farnesoid X receptor. *J Biol Chem*. 2004; 279: 36184-36191.
39. Pols TWH, Puchner HI, Korkmaz T, Vos M, Soeters MR, de Vries CJM, et al. Lithocholic acid controls adaptive immune responses by inhibition of Th1 activation through the Vitamin D receptor. *PLOS ONE*. 2017; 12: e0176715.
40. Unno A, Takada I, Takezawa S, Oishi H, Baba A, et al. TRRAP as a hepatic coactivator of LXR and FXR function. *Biochem Biophys Res Commun*. 2005; 327: 933-938.
41. Viswakarma N, Jia Y, Bai L, Vluggens A, Borensztajn J, et al. Coactivators in PPAR-Regulated Gene Expression. *PPAR Research*. 2010; 21.
42. Wagner M, Halilbasic E, Marschall HU, Zollner G, Fickert P, et al. CAR and PXR agonists stimulate hepatic bile acid and bilirubin detoxification and elimination pathways in mice. *Hepatology*. 2005; 42: 420-430.
43. Wang G, Han T, Wang S, Chen M, Sun Y, et al. Peroxisome Proliferator-Activated Receptor- Prevents Cholesterol Gallstone Formation in C57bl Mice by Regulating Bile Acid Synthesis and Enterohepatic Circulation. *BioMed Research International*. 2018; 12.
44. Wang YX, Lee CH, Tiep S, Yu RT, Ham J, et al. Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell*. 2003; 113: 159-170.